# Evaluation of anti-fibrotic compounds effect in 3D human NASH model using quantitative digital pathology

Radina Kostadinova<sup>1</sup>, Simon Ströbel<sup>1</sup>, Louis Petitjean<sup>2</sup>, Li Chen<sup>2</sup>, Jesús Glaus<sup>1</sup>, Philip Vonschallen<sup>1</sup>, Francisco Verdeguer<sup>1</sup>, Mathieu Petitjean<sup>2</sup> <sup>1</sup>InSphero AG, Schlieren, Switzerland, <sup>2</sup>Pharmanest, Princeton, NJ, USA, Corresponding Author<sup>\*</sup>

## Introduction and Aim

Non-alcoholic steatohepatitis (NASH) is a progressive severe disease characterized by lipid accumulation (steatosis), inflammation (steatohepatitis) and fibrosis in the liver. The development of novel anti-fibrotic therapies has been hindered, in part, by limitations of existing fibrosis analysis techniques of histology samples from in vivo and in vitro preclinical models. FibroNest, a novel Digital Pathology Quantitative AI platform, generates automatic, continuous and direct fibrosis endpoints to quantify fibrosis severity and compound treatment response in clinical NASH samples.

The aim of this study was to establish an algorithm for quantification of fibrosis in an *in* vitro InSight<sup>TM</sup> three-dimensional (3D) human NASH microtissue model using novel digital pathology quantitative single-fiber artificial intelligence (AI) platform FibroNest (PharmaNest). The algorithm was further validated using tool anti-fibrotic compounds such as Alk5 inhibitor and anti-TGF- $\beta$  antibody (AB).

### Method

Using proprietary Akura<sup>™</sup> 96 plate technology for 3D cell culture, we produced 3D human liver microtissues (hLiMTs) using human primary cell types relevant for NASH disease induction and progression: hepatocytes (PHH), Kupffer cells (KC), liver endothelial cells (LSEC) and hepatic stellate cells (HSC). To recapitulate NASH induction *in vitro*, microtissues were exposed for 10 days to defined lipotoxic and inflammatory stimuli, including free fatty acids, high sugar levels, insulin and LPS. Procollagen type I and III secretion were measured using procollagen type I propeptide HTRF assay and procollagen type III propeptide (PIIINP) ELISAs (CisBio), respectively. TIMP-1 and MMP-2 were measured using ProcartaPlex<sup>™</sup> kit and Luminex technology. For FibroNest fibrosis quantification, spheroid FFPE sections were stained with Picro Sirus Red and scanned at 40X. Around 200 principal quantitative fibrosis traits (qFTs) are automatically detected and combined to generate a normalized Phenotypic Composite Fibrosis Score (Ph-FCS). Additional sub-phenotypic fibrosis scores related to collagen fibers, morphometry and architecture are used to further describe the fibrosis phenotypes and its remodeling as fibrosis progress or regresses





Figure 1. InSphero technology and disease modelling. A. Akura 96-well plate technology B. Description of three NASH model treatment conditions (Lean control, NASH induction, NASH induction with treatment) and schedule for endpoint measurement. Compound efficacy studies were performed using NASH induction medium and stimuli to investigate the preventive role of compounds in NASH disease progression



Figure 4. FibroNest fibrosis quantification of anti-fibrotic compounds. A. The Ph-FCS offers a significant detection threshold and dynamic range to evaluate the antifibrotic response of seven clinical compounds treatment arms. p-values are calculated using the Student's T-Test Method

FibroNest algorithm can be used to quantify differences in the fibrosis phenotype in each group and quantify specific effects of each drug (and dose) on the collagen deposition, collagen fibers morphometry and fibrosis architecture.

driven by the compounds, not the Ph-CFS score and method.